



PreHGT: A scalable workflow that screens for horizontal gene transfer within and between kingdoms

Horizontal gene transfer (HGT) is the exchange of DNA between species. It can lead to the acquisition of new gene functions, so finding HGT events can reveal genome novelty. preHGT is a pipeline that uses multiple existing methods to quickly screen for transferred genes.

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Purpose

Horizontal gene transfer (HGT) is the exchange of DNA between an organism and another organism that is not its offspring. It can lead to the rapid acquisition of novel functional traits in the recipient species, leaving distinctive genomic patterns behind in the process. While not all HGT events are maintained in a genome or lead to adaptive

benefit, looking for patterns of HGT across a diverse array of organisms is one way we can survey for functional novelty. Many tools exist for computational discovery of HGT events from genome sequencing data, targeting different genomic patterns and with varying sensitivity, specificity, speed, and scalability. We designed the preHGT pipeline to be a flexible and rapid tool for pre-screening genomes for HGT events. Our goal was to create a pipeline to screen for putative HGT events in as many genomes as are publicly available, or that become available in the future. We wanted an approach that could successfully screen eukaryotic, bacterial, and archaeal genomes and that could screen for transfer events between closely or distantly related species.

The preHGT pipeline uses multiple existing methods for HGT screening and the elimination of false positives. It quickly produces a candidate list of genes that researchers can further investigate with more stringent HGT detection methods, different data modalities, or wet lab experimentation.

We hope this pipeline will be useful to researchers interested in exploring HGT in RefSeq or GenBank genomes.

- This pub is part of the **platform effort**, “[Software: Useful computing at Arcadia](#).” Visit the platform narrative for more background and context.
- The **preHGT pipeline** is available in [this GitHub repository](#).

The context

Adaptation and evolutionary innovation often occur through vertical inheritance and gradual evolutionary processes. Lateral transmission of genomic sequences via HGT is a contrasting evolutionary process that occurs between species instead of from parent to offspring. When genes are transferred, HGT can be a source of rapid functional innovation. Not all HGT events lead to adaptation – some may be neutral, detrimental, or may not be maintained by natural selection and are subsequently lost [1]. Nevertheless, HGT has been the underlying mechanism for many functional adaptations [2][3].

HGT occurs across all domains of life with different frequencies and via many different mechanisms [4][5][6]. In bacteria, HGT most frequently occurs via transduction, conjugation, or transformation. As asexual reproducers with dedicated machinery for HGT, horizontal transfer is one of the most prominent mechanisms for quickly generating genetic diversity. This can catalyze rapid evolution and adaptation to different environmental conditions [3]. However, bacteria also combat HGT by degrading foreign DNA with restriction enzymes and CRISPR [7][8]. Although eukaryotes can undergo HGT through transposable elements, hybridization, and viral transfer, the rate of HGT is relatively low compared to bacteria [5]. This is in part due to structural barriers such as the nucleus that impede the transfer of foreign DNA into the recipient's genome. In sexually reproducing eukaryotes, the frequency of successful horizontal transfer is further reduced because foreign genomic material must reach germ line cells to be transmitted from parent to offspring [9].

Surprisingly, HGT events leave behind similar signatures in recipient genomes independent of the domain of life in which the transfer event occurred. When a gene is transferred, the gene has a different evolutionary history than that of other genes in the recipient's genome. This manifests in different ways depending on how closely related the donor species is to the recipient species. The transferred gene may conserve the function of the gene in the donor genome, may carry a transfer-associated gene annotation, may be abnormally distributed in the species pangenome, or may deviate from species-specific expectations in GC content or other characteristics [10]. The strength of these signals often depends on how much time has passed since the transfer event. Transferred DNA undergoes a process called amelioration, whereby the sequence accumulates mutations over time and becomes less and less distinguishable from the recipient's genome and more and more different from the donor's genome [10]. Other evolutionary processes can further scramble the strength or clarity of a transfer event signature. For example, if many speciation events occurred since the time of the transfer event, it may be difficult to determine whether a horizontal transfer event occurred or if the incongruent evolutionary history is due to other evolutionary processes such as incomplete lineage sorting [11]. If multiple transfer events of the same gene have occurred, or if there have been gene duplications and losses post-speciation, the evolutionary history of a gene may be even more difficult to disentangle. Lastly, convergent evolution and genome contamination can confound HGT discovery by genome sequence analysis as these processes can leave behind similar genomic signatures as bona fide HGT events [12][13][14].

Given this variation, detecting HGT in genome sequence data can be difficult, or at the very least, may require multiple strategies to find different types of transfer events. Luckily, researchers have developed many computational methods to interrogate the genomic signatures left behind in genome sequence data by HGT in different ways (Table 1). These methods fall into two general categories: parametric and phylogenetic [10].

Tool	Category	Taxonomic scope	Event scope	Surrogate
Alien_hunter [15]	Parametric	Bacteria & archaea	Composition	Intervariable composition bias identification through trajectory regression
Alienness [16]	Phylogenetic implicit	All	Kingdom	Mean index score BLAST on sequences
APP [17]	Phylogenetic implicit	Bacteria	Pangenome	Alien Phylogenetic Pathway

Tool	Category	Taxonomic scope	Event scope	Support
				pathogen quarantine disclosure genetic
AnGST [18]	Phylogenetic explicit	All	All	AnGST Genetic Species Collection genetic specimens and discovery uncertainty genetic pathogen critical
AvP [19]	Phylogenetic explicit	All	All	Alien Prevalence Financing host sequestration prevalence mutualism alien and a pathogen and

Tool	Category	Taxonomic scope	Event scope	Sur
				top HG
BLAST2HGT [20]	Phylogenetic implicit	All	Kingdom	Me ind dis ind sc dif fro res
DarkHorse [21]	Phylogenetic implicit	All	Kingdom, sub-kingdom	Me line pro ind BL res
GeneMates [22]	Phylogenetic implicit	Bacteria	Pangenome	Ne ana ge pre abs SN
GIPSy [23]	Parametric, phylogenetic implicit	Bacteria	Composition	Ge Isk Pre So Pre ge

Tool	Category	Taxonomic scope	Event scope	Sur
				isla fea as GC pre mc
HGT-DB [24]	Parametric	Bacteria & archaea	Composition	A d pot eve det de GC coo am usa
HGT-Finder [25]	Phylogenetic implicit	All	Sub- kingdom	Me tra fro res
HGTector [26]	Phylogenetic implicit	All	Sub- kingdom	Me like HG bet anc dis fro res

Tool	Category	Taxonomic scope	Event scope	Surveillance
HGTphiloDetect [27]	Phylogenetic implicit, phylogenetic explicit	All	All	Metagenomic, individual, outbreak, BLAST, reservoir, follow-up, phylogenetic information, initial characterization
HGTree [28]	Phylogenetic explicit	Bacteria & archaea	All	Adaptive potential, evolution, usage, reconstruction
Islander [29]	Parametric	Bacteria	Bacteria	Taxonomic identification of the island
IslandHunter [30]	Parametric	Bacteria	Composition	Presence of genomic islands, features as GC content, presence of mobile elements

Tool	Category	Taxonomic scope	Event scope	Sur
IslandPath-DIMOB [31]	Parametric	Bacteria	Composition	Pre ge isl di co an of ge
IslandPick [32]	Phylogenetic implicit	Bacteria	Species, Strain	Pre ge isl co clo ge
IslandViewer4 [33]	Parametric, phylogenetic implicit	Bacteria & archaea	See other tools	Int Isl Isl DIM HM Isl
Near HGT [34]	Phylogenetic implicit	Bacteria	Species, Strain	Me syn and rel mu co

Tool	Category	Taxonomic scope	Event scope	Sur
PGAP-X [35]	Phylogenetic implicit	Bacteria	Pangenome	Pal An Pip Pal gel abs
RANGER-DTL [36]	Phylogenetic explicit	All	All	Ra of Ev Re DT Re gel spe to du tra los
RecentHGT [37]	Phylogenetic implicit	Bacteria & archaea	Species, Strain	Ex ma alg glo sec ali
RIATA-HGT [38]	Phylogenetic explicit	All	All	Ide inc be

Tool	Category	Taxonomic scope	Event scope	Surveillance scope
				tree structure
SIB [39]	Parametric	Bacteria & archaea	Species, Strain	Sequence information, Bionomics, Significance from occurrence data, trajectory
ShadowCaster [40]	Parametric, phylogenetic explicit	Bacteria & archaea	Composition	Use of vector on community composition, feature identification, calculation of the relative abundance of orthologs, significance in taxonomic distribution
SigHunt	Parametric	Eukaryotes	Composition	Slit of 4

Tool	Category	Taxonomic scope	Event scope	Sur
[41]				fre
SIGI-HMM [42]	Parametric	Bacteria & archaea	Composition	Pre ge isl co co bia hid mc
T-REX [43]	Phylogenetic explicit	All	All	Tre se Re Evo Inc ph tre
TF-IDF [44]	Parametric	Bacteria & archaea	Species, Strain	Ter fre inv do fre ide uni se fea

Table 1. Non-exhaustive list of computational tools for HGT discovery.

Composition: Composition different from acceptor genome.

Pangenome: Any set of organisms one can reasonably build a pangenome from (clade, species, genus).

Kingdom: Cross-kingdom detection, usually by user-defined definition of ingroup and outgroup.

Sub-kingdom: Any taxonomic level lower than kingdom and higher than species or strain, usually with decreasing accuracy at higher taxonomic resolution.

Parametric methods analyze the genome of interest to identify regions that deviate from species-specific expectations in GC content, codon usage, amino acid usage, k-mer frequencies, gene annotations, or other characteristics [10]. These methods are fast, but natural differences in genome uniformity can lead to over-prediction and they are often limited to recent transfer events for which amelioration of transferred DNA is limited [10]. Parametric approaches can also be biased by gene length [45][46], so they may be difficult or impossible to use on genes, which vary in size, as opposed to sliding windows across the genome, which are a consistent length.

Phylogenetic methods detect inconsistencies between gene and species evolution [10]. This category can be further divided into explicit and implicit methods. Explicit methods test alternative evolutionary scenarios using tree-based analysis, while implicit methods rely on implied phylogenetic relationships derived from comparative genomic approaches. Gene-by-gene explicit phylogenetic methods are the gold standard in horizontal gene transfer detection [10][47]. The most robust of these approaches works by formally reconciling gene family tree topologies (where each tip is a protein sequence belonging to a species) with the species tree topology (each tip is a species) under explicit Maximum Likelihood inference for models of gene family duplication, transfer, and loss [48][49]. These methods identify candidate ancestral HGT events while accounting for the confounding impacts of gene duplication and loss on these inferences. Although powerful, these methods require that gene homology is already known and that gene family trees of these homologous sequences have already been inferred. Consequently, these methods are typically ideally suited for focused application to a set of gene families of special interest and thus are less computationally tractable to apply at scale than other HGT prediction methods.

Without *a priori* knowledge about the donor and recipient genomes for horizontally transferred genetic material, it becomes necessary to sample in a taxonomically broad and unbiased manner. In this respect, implicit phylogenetic methods are particularly well suited to hypothesis-free discovery of HGT events, as they scale more readily to hundreds of genomes than do explicit methods. Implicit methods rely on patterns that

correlate with evolutionary history to infer HGT. For example, you can use BLAST to identify homologous genes with different taxonomic labels than the query gene, which can be analyzed to find patterns consistent with HGT [19][25][50][51][52]. Similarly, you can use the pangenome – the full complement of genes shared between a set of closely related organisms – to investigate HGT by determining the presence or absence of genes across all genomes [53][54].

Across the HGT literature and tool space, including both parametric and phylogenetic methods, genome contamination is often underappreciated. Contaminant sequences in genomes can look like HGT events. This has led to rebuttals [14][55] against high-profile papers [56][57] that claimed detection of high fractions of horizontally transferred genes, and may more generally impact the biological interpretation of HGT predictions. At least 0.54% of genomes in GenBank and 0.34% in RefSeq are contaminated [58]. While some methods incorporate careful contamination checks [19], others rely on filtering heuristics [16] or omit them entirely.

The problem

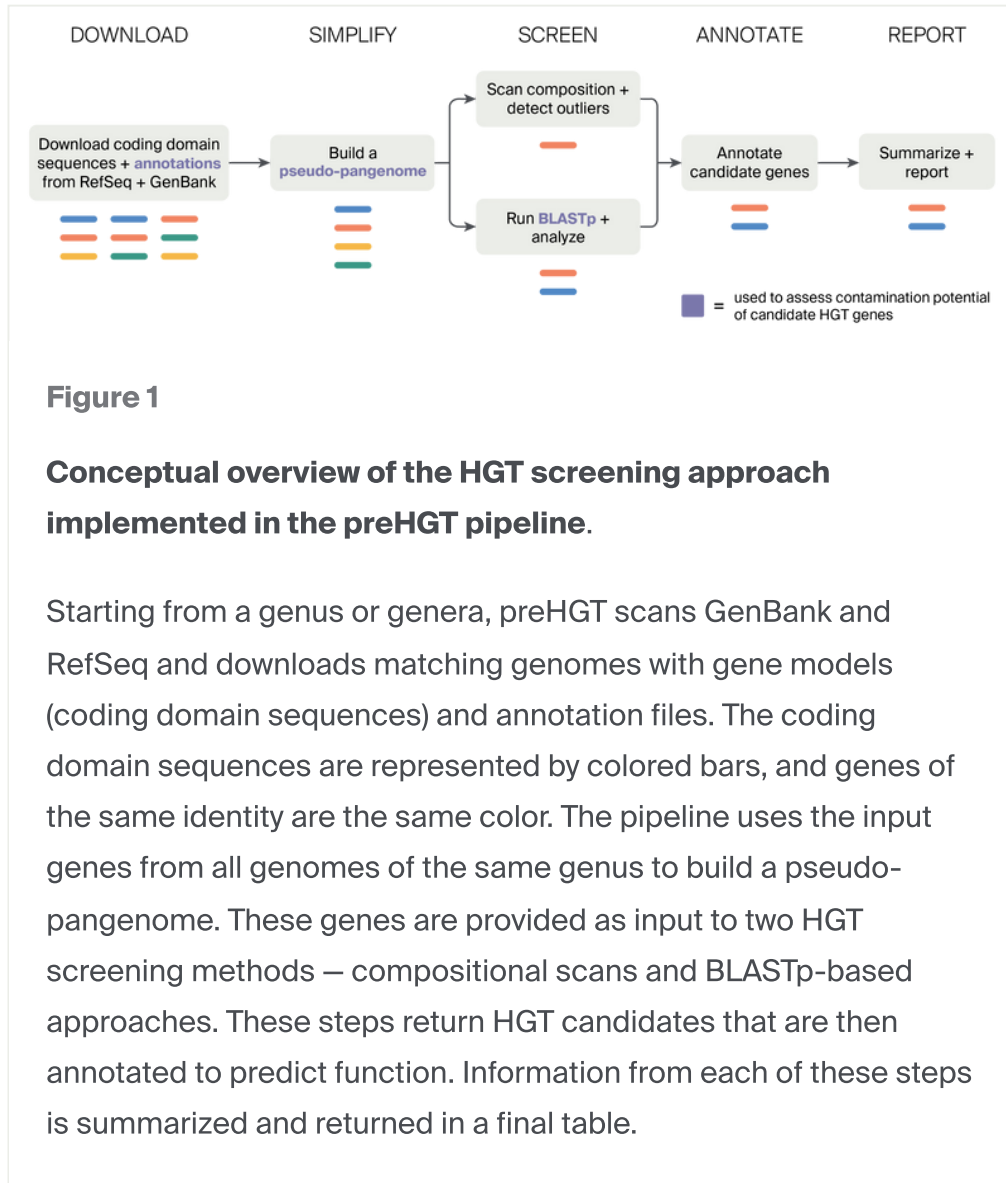
We sought a scalable computational approach for predicting HGT candidate genes. We wanted the pipeline to be able to screen for HGT events across the tree of life and across taxonomic scopes (from family- to kingdom-level transfers), and to assess the likelihood that a candidate transfer event was instead the result of genome contamination.

As other projects at Arcadia are developing explicit phylogenetic methods for the inference of gene family evolution, we sought a solution that we could use upstream of this tool to produce candidate species lists for further validation, and tried to avoid using trees so as not to duplicate efforts.

Our solution

We built a pipeline that we're calling "preHGT" to quickly find *preliminary HGT* candidates in genomes with gene predictions. Our approach blends parametric and phylogenetic implicit methods to generate a list of candidate genes that may have been horizontally transferred ([Figure 1](#)). The preHGT pipeline uses compositional

scans, pangenome inference, and BLAST-based searches. It combines information from these approaches, as well as annotation information, to highlight candidate genes that are more likely to be contamination than HGT. By implementing multiple HGT screens in one pipeline, we aimed to combine approaches that target different signatures of HGT, to provide a more comprehensive HGT screening strategy.



As we were designing the pipeline, we were concerned about overall run times, especially given that BLAST searches can be computationally expensive. We implemented clustering heuristics at two key places to keep the pipeline fast. First, we clustered the genes in input genomes to reduce the number of genes we investigated for HGT potential. Given our eventual goal of running this pipeline on all publicly available genomes, we wanted to assess the potential for HGT in redundant genes only once. We did this by clustering genes in closely related genomes – those of the

same genus – prior to screening for HGT. Second, we clustered the NCBI BLAST non-redundant protein database, reducing its size by over half, to increase the speed of BLAST searches [59].

One of the reasons we were particularly excited to include BLAST in our pipeline was to take advantage of a rich literature of BLAST-based HGT predictor indices (Table 1, Table 2). Many creative and insightful HGT screening methods exist, each with its own strengths. However, these methods are contained in different tools. Since BLAST is the most expensive computational step of our pipeline and none of the methods rely on a clustered BLAST database, we re-implemented them in the preHGT workflow. This consolidation allows HGT screening using a single tool and a single BLAST run (Table 2).

We implemented the pipeline as both a Snakemake [60] and a Nextflow [61] workflow, with software environments controlled by [conda](#) or Docker. The modular nature of the workflow will allow us to incorporate additional methods over time.

The preHGT pipeline does not implement any new algorithms for HGT candidate screening. However, the pipeline contributes to this space by:

1. Combining multiple existing HGT screening algorithms in one pipeline.
2. Using pangenome inference on eukaryotic genomes to inform a gene's contamination potential and phyletic distribution, and to reduce compute required to run the pipeline.
3. Reducing the BLAST database size by clustering similar proteins, thereby reducing compute required to run the pipeline and diversifying taxonomic lineages represented in top hits.
4. Providing multiple information sources to help assess an HGT candidate's contamination potential.

The resource

The **preHGT Snakemake and Nextflow workflows** are available at [this GitHub repository](#) (DOI: [10.5281/zenodo.8169269](#)).

Below we provide an overview of each step in the preHGT pipeline ([Figure 2](#)).

1. **Retrieving gene sequences and annotation files.** The pipeline begins with the user providing a genus or genera of interest in a TSV file. The pipeline then scans GenBank [62] and RefSeq [63] for matching genomes and downloads gene models and genome annotation files using `ncbi-genome-download`. When a genome is available in both GenBank and RefSeq, only the RefSeq version is retained.
2. **Building a pseudo-pangenome.** For each genus, the pipeline then combines genes into a pseudo-pangenome by clustering the nucleotide sequences at 90% length and identity using `mmseqs easy-cluster` [64]. For each cluster, MMSeq2 selects a single representative sequence by retaining the sequence with the most alignments. The clustered nucleotide sequences are then translated into amino acid sequences using `EMBOSS transeq` [65].

We refer to this as a pseudo-pangenome, and not a pangenome, because we empirically cluster genes based on sequence similarity and not by constructing orthologous groups or by considering the evolutionary history of each sequence [66][67]. We recognize that while this may collapse functionally different paralogs, it is unlikely to obscure patterns of HGT from distant donor genomes; paralogous genes share a common ancestor, so while they may serve different purposes for the organism, at >90% identity only one copy of the gene needs to be screened for HGT potential. Using a pseudo-pangenome is useful in two ways for the pipeline. First, it reduces the number of genes that are investigated which reduces run times. Second, it provides metadata about the gene. Singletons are more likely to be contaminants, and genes that are only present in a subset of genomes may have interesting evolutionary histories (e.g. gene loss).

3. **Screening for HGT candidates.** Using the genes in the pseudo-pangenome, the preHGT pipeline then uses two approaches to screen for HGT candidates.
 - **Compositional scan.** The first approach uses relative amino acid usage to detect proteins with outlying composition. It measures relative amino acid usage using the `EMBOSS pepstats` function [65], produces a distance matrix with the base R function `dist()`, and hierarchically clusters the distance matrix with `fastcluster`'s `hclust` [68]. It detects outliers by cutting the resultant tree with `height/1.5` and retaining any cluster that contains fewer than 0.1% of the pseudo-pangenome size.

Relative amino acid usage is the frequency that each amino acid is used in each gene, normalized by the total number of amino acids in that gene. For example, if alanine is used 27 times in a protein that is 100 amino acids long, the relative usage would be 27%. Relative amino acid usage is generally conserved across a genome and reflects an organism's environment [69]. We tried many compositional metrics such as tetranucleotide frequency, GC content, and codon usage. However, we found that outlying proteins were driven by abnormal length for all metrics other than relative amino acid usage.

Given that this is a reference-free approach, genes returned by this screening method do not have accompanying donor species predictions, which makes interpretation more challenging. Aberrant relative amino acid usage can also arise from mechanisms other than HGT and this method does not distinguish between potential sources.

- **BLASTp scan.** The second approach uses BLASTp to identify homologous proteins. All genes in the pseudo-pangenome are BLASTed against a clustered version of NCBI's nr database (90% length, 90% identity) [59] using DIAMOND `blastp` [70]. The pipeline then adds lineage information to the BLASTp search using `dplyr`, `dbplyr`, and `RSQLite` [71]. It scans these results for signatures of transfer events using multiple, previously published algorithms (Table 2) [19][25][50][51][52].

One modification we made throughout is using length-corrected bit scores output by DIAMOND `blastp` instead of raw bit scores. Bit scores are sensitive to gene length, so using corrected bit scores reduces biases associated with gene length in HGT screening [72].

The choice of database will dramatically impact the results produced by this screen. We chose to use a clustered version of the NCBI nr database [59] both to make the BLASTp step faster and to ensure the results contain a variety of taxonomic lineages in cases where many near and distant homologs exist. Using this database, combined with our methods of choice (Table 2), the preHGT pipeline screens for HGT events that occur in seven domains of NCBI's taxonomy: bacteria, archaea, fungi, plants, metazoa, other eukaryotes, and viruses ("kingdom" taxonomic resolution). It will also screen for HGT events

between lineages that are in the same domain as the query genus but are different up to the family level from that genus (“sub-kingdom” taxonomic resolution).

Index	Tool	Taxonomic resolution	Data used	Calculated by
Aggregate hit support ⁺ [19]	AvP	Kingdom	All bit scores	Subtracting the sum of normalized bit scores in the donor group from the sum of normalized bit scores in the acceptor group.
Alien index [50]	NA	Kingdom	Minimum e-value	Subtracting the transformed e-value of the best donor hit from the transformed e-value of the best non-self acceptor hit.
HGT score [51]	NA	Kingdom	Maximum bit score	Subtracting the best non-self acceptor hit bit score from the best donor hit bit score and normalizing this value.
Donor distribution index [52]	NA	Kingdom	Number of hits per kingdom	Measuring the dispersion query homologs across groups by determining the number of hits per kingdom against the total number of possible kingdoms.
Gini coefficient	NA	Kingdom	Number of hits per kingdom	Measuring inequality among values of a distribution, where values are the number of BLAST hits observed for each kingdom.
Entropy	NA	Kingdom	Number of hits per kingdom	Measuring disorder among values, where values are the number of BLAST hits observed for each kingdom.
Transfer index [25]	HGT-Finder	Kingdom, Sub-	All bit scores	Considering taxonomic distances between

Index	Tool	Taxonomic resolution	Data used	Calculated by
		kingdom		query and hit, bit score ratios, and rank and total number of BLAST hits.

Table 2. Algorithms that parse BLASTp results to predict HGT candidates.

*NA: Not applicable.

⁺ Aggregate hit support is calculated by subtracting the sum of all normalized BLAST bit scores for all hits in an in-group from an out-group. We use a different normalization equation than the original method, which leads to different results.

1. **Annotation.** We then annotate the HGT candidates. For each candidate HGT amino acid sequence, we use two different approaches for ortholog annotation. First, the pipeline uses KofamScan for KEGG ortholog annotation [73]. Next, the pipeline uses HMMER3 `hmmsearch` to assign annotations to HGT candidates. `hmmsearch` compares each HGT candidate sequence against hidden Markov models (HMMs) of proteins in a database. We built a custom HMM database to target specific annotations of interest. The HMM database currently contains Virus Orthologous Groups from VOGDB and biosynthetic genes, and can be extended in the future to meet user annotation interests.
2. **Reporting.** The last step combines all information that the pipeline has produced and outputs the results in a TSV file. The results include the GenBank protein identifier for the HGT candidate, BLAST and relative amino acid usage scores, pangenome information, gene and ortholog annotations, and contextualizing information about the gene such as position in the contiguous sequence.

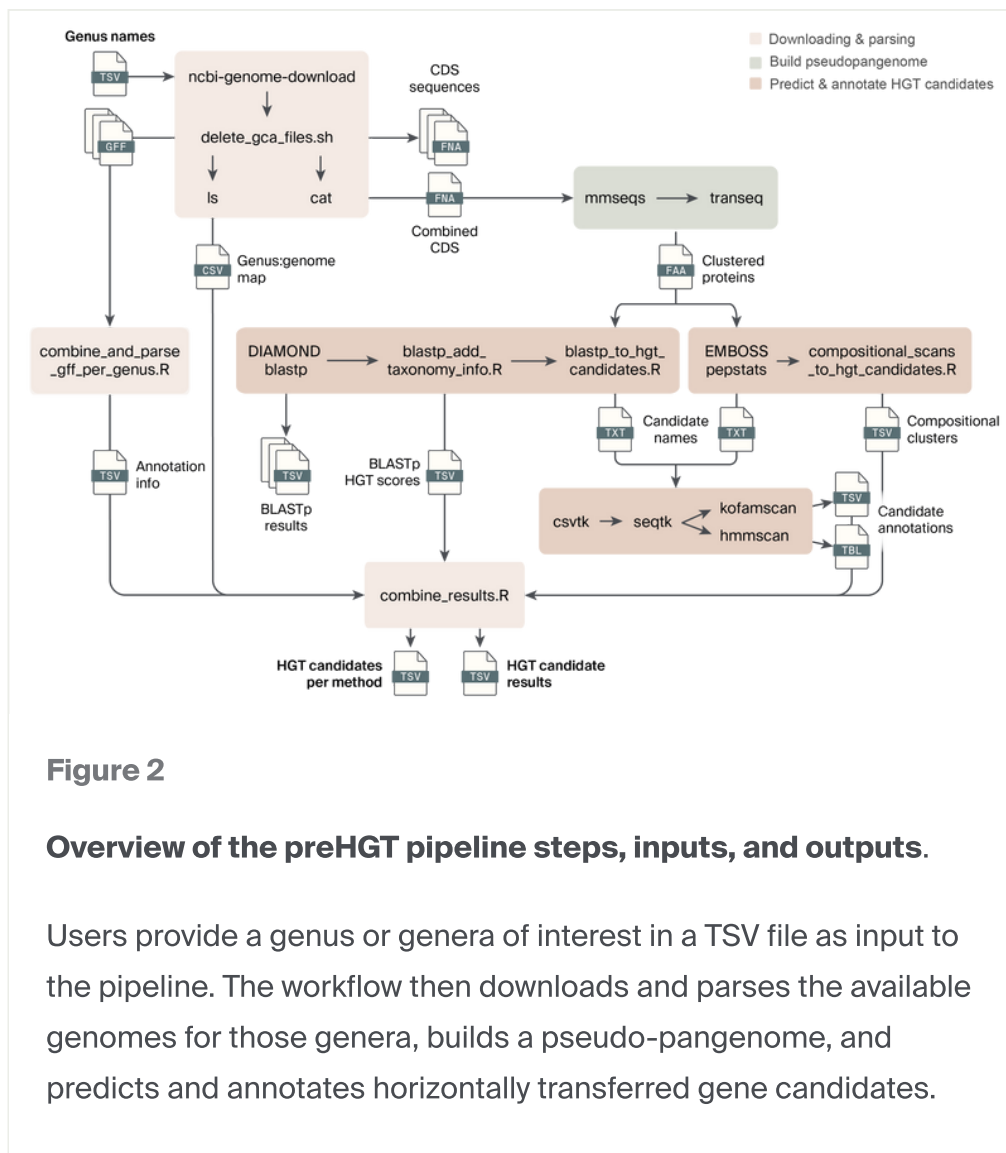


Figure 2

Overview of the preHGT pipeline steps, inputs, and outputs.

Users provide a genus or genera of interest in a TSV file as input to the pipeline. The workflow then downloads and parses the available genomes for those genera, builds a pseudo-pangenome, and predicts and annotates horizontally transferred gene candidates.

Types of HGT events that the pipeline screens for

While we tried to create a fast and generalized pipeline, preHGT is better at detecting some patterns of HGT than others. The preHGT pipeline screens for HGT events where the donor and recipient differ in taxonomy at the family level or above. It is most likely more accurate when the transfer events occur between more distantly related organisms and where the recipient gene retains homology to the gene in the donor genome. We anticipate the primary use of this approach will be to identify candidate transfer events and donor and recipient groups to which more granular approaches can be applied to better disentangle the evolutionary history of the gene.

The parametric approach we implemented screens for genes with outlying relative amino acid usage compared to the rest of the genes in the genome. This requires that the donor and acceptor species differ in amino acid composition, and that these differences persist in the transferred genes, a scenario that is most typical of recent transfer events among evolutionarily divergent species.

The BLAST-based implicit phylogenetic approaches we implemented screen for genes that exhibit a greater degree of sequence similarity among designated taxonomic outgroups than within ingroups. In the original tools and papers in which these algorithms were generated, the authors implemented or validated their approaches at specific taxonomic levels that the preHGT pipeline adheres to (Table 2). Some are designed to screen for cross-kingdom transfer events, while others can screen for sub-kingdom-level events. However, because the chance of spurious inference of homology increases among more closely related species, results should be more carefully scrutinized at lower taxonomic levels (e.g. order, family). Homology detection also becomes increasingly difficult at larger taxonomic distances, so highly diverged homologs may be missed by the pipeline.

Additional considerations and caveats

How we deal with contamination and other sources of false positives

HGT screens often return many false positives [56][57]. We used contextualizing information about HGT candidates to reduce the number of false positives reported by the pipeline.

Contamination is the biggest source of false positives in BLAST-based HGT screening algorithms. Many genomes in GenBank and RefSeq are contaminated [58].

Contamination arises from impure sampling, contaminated reagents, lab cross-contamination, sequencing artifacts, or reference database errors [74]. To combat the presence of contamination, we incorporated multiple corroborating lines of evidence to assess whether contamination is more likely than HGT. First, we determine the length of the contiguous sequence within which the candidate gene is found. Short contiguous sequences are more likely to be contaminants [58][75]. Next, we determine how many genes are in the candidate gene's cluster from our pseudo-

pangenome approach. Depending on the contamination source, it is unlikely that the same contamination will occur in multiple genomes [76]. Therefore, if a homolog is present in multiple genomes, it is less likely to be a contaminant. Lastly, for BLAST-based results, we assess the percent identity between the donor and acceptor genes. Amelioration deteriorates sequence identity after a transfer event [10], so the more similar two genes are, the more likely similarity is driven by contamination. Many methods use a cutoff of 70%–80% identity for contamination [16][77], but we instead weigh this against other corroborating information.

In the future, we hope to further contextualize contamination potential against the general contamination score for the acceptor genome. The more contamination a genome contains, the more likely a candidate is to be a contaminant itself.

BLAST-based methods may also generate false positives arising from alignment errors or alignment due to sequence similarity that does not arise from shared ancestry, such as from convergent evolution or random chance. Alignment errors from short or low-complexity sequences or from short, highly conserved domains may give the appearance of a horizontal transfer event. To protect against this, we filter corrected bit scores to those greater than 100, or, to rescue true homologs that are very divergent, with a query coverage of greater than 70%. We also provide gene annotations from multiple annotation sources to highlight hits that might be ultra-conserved, such as those from ribosomal proteins. Over time, we hope to curate a list of genes that the preHGT pipeline frequently detects as false positives and to develop a strategy to filter them out.

Verifying bona fide HGT requires work beyond preHGT

The preHGT pipeline provides a list of candidate HGT events. These candidates need to be carefully scrutinized to determine whether they are biologically interesting and whether they are more or less likely to be false positives. We built preHGT as a generalized precursor to more in-depth HGT analysis (Figure 3). We envision that preHGT can inform genome selection for comprehensive explicit phylogenetic inference, which can help disentangle alternate evolutionary trajectories, or highlight when not enough information is available to support HGT inference.

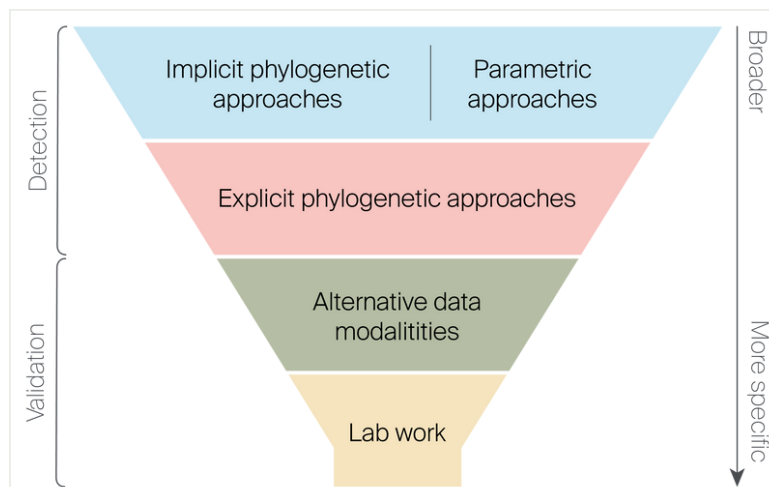


Figure 3

Funnel of methods for HGT screening and validation.

Implicit phylogenetic and parametric approaches are fast, generalized methods for screening for HGT in genes or genomes, but these methods are prone to false positives. Explicit phylogenetic methods can help eliminate some false positives or determine when there is not enough evidence to support HGT candidacy.

After these computational approaches, validation requires additional methods. Alternative data modalities like transcriptome sequencing or laboratory experiments like FISH or PCR can provide additional evidence in support of HGT. While powerful, these methods require curated information about the donor and acceptor genomes and the candidate genes and thus can usually only be used after initial exploration.

After phylogenetic analysis, more analysis is still required to reject the null hypothesis that no transfer event occurred. The appropriate experiments for this will depend on the HGT candidate event itself. For example, if a bacterial gene has been transferred into a eukaryotic genome, it may be appropriate to interrogate the candidate gene for

the presence of introns, or if transcriptome information is available, for the presence of transcription- and eukaryotic-specific RNA modifications such as 5' caps or Kozak sequences. In the lab, PCR, FISH, or Southern blots may confirm the presence of the sequence in the genome of interest, while Western blot or mass spectrometry can confirm that the gene is transcribed and translated into a protein.

Limitations of the preHGT pipeline

Given our approach, we have identified multiple shortcomings. The most conspicuous limitation is our focus on genes. The preHGT pipeline can only scan genomes with gene models. We elected not to implement genome annotation as an early step in the pipeline given that annotation procedures differ for eukaryotic versus bacterial and archaeal genomes, and that eukaryotic genome annotation remains a challenging problem from the genome alone [78][79]. This limits the preHGT pipeline to those genomes with gene models (approximately 21%) and creates blind spots for HGT detection across the tree of life. Of 56 eukaryotic phyla with genomes, only 45 have at least one genome with gene models. Similarly, by treating genes in their entirety as the unit that is horizontally transferred, we are unlikely to detect genes for which only a nested region of the coding sequence was horizontally transferred.

There are also limitations born out of our decision to use composition or BLAST-based HGT screening methods. First, these methods require that the gene has not ameliorated to the composition of the acceptor genome or that it maintains detectable homology to the donor genome. This may limit our detection of ancient HGT events. Second, these methods will be less sensitive to HGT events that occur between closely related organisms. Third, since BLAST-based approaches rely on taxonomies, there are risks since taxonomies may be wrong and since they do not account for branch lengths in the relatedness of species. Lastly, false positives may arise from alignment between short or low-complexity sequences or from natural sequence similarity such as what might arise from convergent evolution or from highly conserved gene sequences. To combat both cases, we have implemented filtering criteria to help eliminate these issues.

Lastly, we did not integrate an explicit phylogenetic approach to better resolve the evolutionary histories of HGT candidates. We elected to forgo this step because another team at Arcadia is developing a tree-based workflow. We are currently

experimenting with how to facilitate handoff between the two tools to rapidly enable this next step in validation.

Additional methods

We used ChatGPT to add comments to our code and suggest wording ideas. We also used ChatGPT to add comments to external code to help us better understand how it worked when trying to implement some existing tools in another language.

Key takeaways

- preHGT is a scalable pipeline that screens for potential HGT events in genomes with gene models across the tree of life and taxonomic scales.
- The pipeline leverages compositional and BLASTp scans, pangenome inference, annotation, and reporting techniques to provide comprehensive results.
- Multiple checks and filters defend against false positives, including contamination detection and sequence alignment artifact filtering.
- The pipeline is implemented in both Snakemake and Nextflow. Its modular design means it's easily extensible to incorporate more methods in the future.
- preHGT aims to identify HGT events that users further investigate with other approaches such as tree-based ones.

Next steps

Our follow-up plans include:

1. **Eukaryotic HGT prediction:** We plan to run the pipeline on all eukaryotic genomes in GenBank and RefSeq that have gene models and to make the results available.
2. **Building a user interface for results exploration:** We plan to build a simple user interface to explore results produced by the pipeline. Exploration modes will allow users to dive into gene transfer events by donor or acceptor taxonomy, predicted

functions of genes involved, or by strength of result, and to visualize the results in their genomic context.

3. **Adapting the pipeline to take transcriptome assemblies as input:** We plan to extend the pipeline to run on assembled transcriptomes by incorporating upstream gene prediction rules. We will then run the pipeline on the transcriptomes in the NCBI Transcriptome Shotgun Assembly database and make the results publicly available.
4. **Integrating new algorithms for HGT screening:** Other algorithms exist for the interpretation of BLASTp results. We plan to integrate those from other tools into this pipeline in the future.

We welcome feedback on the user experience, the results we include, or additional algorithms or metrics that would be helpful to incorporate.

References

- 1 Soucy SM, Huang J, Gogarten JP. (2015). Horizontal gene transfer: building the web of life. <https://doi.org/10.1038/nrg3962>
- 2 Husnik F, McCutcheon JP. (2017). Functional horizontal gene transfer from bacteria to eukaryotes. <https://doi.org/10.1038/nrmicro.2017.137>
- 3 Arnold BJ, Huang I-T, Hanage WP. (2021). Horizontal gene transfer and adaptive evolution in bacteria. <https://doi.org/10.1038/s41579-021-00650-4>
- 4 Zaneveld JR, Nemergut DR, Knight R. (2008). Are all horizontal gene transfers created equal? Prospects for mechanism-based studies of HGT patterns. <https://doi.org/10.1099/mic.0.2007/011833-0>
- 5 Van Etten J, Bhattacharya D. (2020). Horizontal Gene Transfer in Eukaryotes: Not if, but How Much? <https://doi.org/10.1016/j.tig.2020.08.006>
- 6 Wagner A, Whitaker RJ, Krause DJ, Heilers J-H, van Wolferen M, van der Does C, Albers S-V. (2017). Mechanisms of gene flow in archaea. <https://doi.org/10.1038/nrmicro.2017.41>

- 7 Corvaglia AR, François P, Hernandez D, Perron K, Linder P, Schrenzel J. (2010). A type III-like restriction endonuclease functions as a major barrier to horizontal gene transfer in clinical *Staphylococcus aureus* strains. <https://doi.org/10.1073/pnas.1000489107>
- 8 Marraffini LA, Sontheimer EJ. (2008). CRISPR Interference Limits Horizontal Gene Transfer in Staphylococci by Targeting DNA. <https://doi.org/10.1126/science.1165771>
- 9 Gabaldón T. (2020). Patterns and impacts of nonvertical evolution in eukaryotes: a paradigm shift. <https://doi.org/10.1111/nyas.14471>
- 10 Ravenhall M, Škunca N, Lassalle F, Dessimoz C. (2015). Inferring Horizontal Gene Transfer. <https://doi.org/10.1371/journal.pcbi.1004095>
- 11 Knowles LL, Huang H, Sukumaran J, Smith SA. (2018). A matter of phylogenetic scale: Distinguishing incomplete lineage sorting from lateral gene transfer as the cause of gene tree discord in recent versus deep diversification histories. <https://doi.org/10.1002/ajb2.1064>
- 12 Copley SD, Dhillon JK. (2002). Lateral gene transfer and parallel evolution in the history of glutathione biosynthesis genes. <https://doi.org/10.1186/gb-2002-3-5-research0025>
- 13 Cote-L'Heureux A, Maurer-Alcalá XX, Katz LA. (2022). Old genes in new places: A taxon-rich analysis of interdomain lateral gene transfer events. <https://doi.org/10.1371/journal.pgen.1010239>
- 14 Koutsovoulos G, Kumar S, Laetsch DR, Stevens L, Daub J, Conlon C, Maroon H, Thomas F, Aboobaker AA, Blaxter M. (2016). No evidence for extensive horizontal gene transfer in the genome of the tardigrade *Hypsibius dujardini*. <https://doi.org/10.1073/pnas.1600338113>
- 15 Vernikos GS, Parkhill J. (2006). Interpolated variable order motifs for identification of horizontally acquired DNA: revisiting the *Salmonella* pathogenicity islands. <https://doi.org/10.1093/bioinformatics/btl369>
- 16 Rancurel C, Legrand L, Danchin E. (2017). Alienness: Rapid Detection of Candidate Horizontal Gene Transfers across the Tree of Life. <https://doi.org/10.3390/genes8100248>
- 17 Sengupta S, Azad RK. (2023). Leveraging comparative genomics to uncover alien genes in bacterial genomes. <https://doi.org/10.1099/mgen.0.000939>
- 18 David LA, Alm EJ. (2010). Rapid evolutionary innovation during an Archaeal genetic expansion. <https://doi.org/10.1038/nature09649>

- 19 Koutsovoulos GD, Granjeon Noriot S, Bailly-Bechet M, Danchin EGJ, Rancurel C. (2022). AvP: A software package for automatic phylogenetic detection of candidate horizontal gene transfers. <https://doi.org/10.1371/journal.pcbi.1010686>
- 20 Li M, Zhao J, Tang N, Sun H, Huang J. (2018). Horizontal Gene Transfer From Bacteria and Plants to the Arbuscular Mycorrhizal Fungus *Rhizophagus irregularis*. <https://doi.org/10.3389/fpls.2018.00701>
- 21 Podell S, Gaasterland T. (2007). DarkHorse: a method for genome-wide prediction of horizontal gene transfer. <https://doi.org/10.1186/gb-2007-8-2-r16>
- 22 Wan Y, Wick RR, Zobel J, Ingle DJ, Inouye M, Holt KE. (2020). GeneMates: an R package for detecting horizontal gene co-transfer between bacteria using gene-gene associations controlled for population structure. <https://doi.org/10.1186/s12864-020-07019-6>
- 23 Soares SC, Geyik H, Ramos RTJ, de Sá PHCG, Barbosa EGV, Baumbach J, Figueiredo HCP, Miyoshi A, Tauch A, Silva A, Azevedo V. (2016). GIPSy: Genomic island prediction software. <https://doi.org/10.1016/j.jbiotec.2015.09.008>
- 24 Garcia-Vallve S. (2003). HGT-DB: a database of putative horizontally transferred genes in prokaryotic complete genomes. <https://doi.org/10.1093/nar/gkg004>
- 25 Nguyen M, Ekstrom A, Li X, Yin Y. (2015). HGT-Finder: A New Tool for Horizontal Gene Transfer Finding and Application to *Aspergillus* genomes. <https://doi.org/10.3390/toxins7104035>
- 26 Zhu Q, Kosoy M, Dittmar K. (2014). HGTector: an automated method facilitating genome-wide discovery of putative horizontal gene transfers. <https://doi.org/10.1186/1471-2164-15-717>
- 27 Yuan L, Lu H, Li F, Nielsen J, Kerkhoven EJ. (2023). HGTphyloDetect: facilitating the identification and phylogenetic analysis of horizontal gene transfer. <https://doi.org/10.1093/bib/bbad035>
- 28 Choi Y, Ahn S, Park M, Lee S, Cho S, Kim H. (2022). HGTree v2.0: a comprehensive database update for horizontal gene transfer (HGT) events detected by the tree-reconciliation method. <https://doi.org/10.1093/nar/gkac929>
- 29 Hudson CM, Lau BY, Williams KP. (2014). Islander: a database of precisely mapped genomic islands in tRNA and tmRNA genes. <https://doi.org/10.1093/nar/gku1072>
- 30 Baichoo S, Goodur H, Ramtohul V. (2014). IslandHunter – A Java-based GI detection software. <https://doi.org/10.7287/peerj.preprints.716v1>

- 31 Bertelli C, Brinkman FSL. (2018). Improved genomic island predictions with IslandPath-DIMOB. <https://doi.org/10.1093/bioinformatics/bty095>
- 32 Langille MG, Hsiao WW, Brinkman FS. (2008). Evaluation of genomic island predictors using a comparative genomics approach. <https://doi.org/10.1186/1471-2105-9-329>
- 33 Bertelli C, Laird MR, Williams KP, Lau BY, Hoad G, Winsor GL, Brinkman FS. (2017). IslandViewer 4: expanded prediction of genomic islands for larger-scale datasets. <https://doi.org/10.1093/nar/gkx343>
- 34 Adato O, Ninyo N, Gophna U, Snir S. (2015). Detecting Horizontal Gene Transfer between Closely Related Taxa. <https://doi.org/10.1371/journal.pcbi.1004408>
- 35 Zhao Y, Sun C, Zhao D, Zhang Y, You Y, Jia X, Yang J, Wang L, Wang J, Fu H, Kang Y, Chen F, Yu J, Wu J, Xiao J. (2018). PGAP-X: extension on pan-genome analysis pipeline. <https://doi.org/10.1186/s12864-017-4337-7>
- 36 Bansal MS, Kellis M, Kordi M, Kundu S. (2018). RANGER-DTL 2.0: rigorous reconstruction of gene-family evolution by duplication, transfer and loss. <https://doi.org/10.1093/bioinformatics/bty314>
- 37 Li X, Tong W, Wang L, Rahman SUr, Wei G, Tao S. (2018). A Novel Strategy for Detecting Recent Horizontal Gene Transfer and Its Application to Rhizobium Strains. <https://doi.org/10.3389/fmicb.2018.00973>
- 38 Nakhleh L, Ruths D, Wang L-S. (2005). RIATA-HGT: A Fast and Accurate Heuristic for Reconstructing Horizontal Gene Transfer. https://doi.org/10.1007/11533719_11
- 39 Narechania A, Bobo D, DeSalle R, Mathema B, Kreiswirth B, Planet PJ. (2021). What do we gain when tolerating loss? The information bottleneck wrings out recombination. <https://doi.org/10.1101/2021.08.27.457981>
- 40 Sánchez-Soto D, Agüero-Chapin G, Armijos-Jaramillo V, Perez-Castillo Y, Tejera E, Antunes A, Sánchez-Rodríguez A. (2020). ShadowCaster: Compositional Methods under the Shadow of Phylogenetic Models to Detect Horizontal Gene Transfers in Prokaryotes. <https://doi.org/10.3390/genes11070756>
- 41 Jaron KS, Moravec JC, Martínková N. (2013). Sig Hunt: horizontal gene transfer finder optimized for eukaryotic genomes. <https://doi.org/10.1093/bioinformatics/btt727>
- 42 Waack S, Keller O, Asper R, Brodag T, Damm C, Fricke WF, Surovcik K, Meinicke P, Merkl R. (2006). Score-based prediction of genomic islands in prokaryotic genomes using hidden Markov models. <https://doi.org/10.1186/1471-2105-7-142>

- 43 Boc A, Diallo AB, Makarenkov V. (2012). T-REX: a web server for inferring, validating and visualizing phylogenetic trees and networks. <https://doi.org/10.1093/nar/gks485>
- 44 Cong Y, Chan Y, Ragan MA. (2016). A novel alignment-free method for detection of lateral genetic transfer based on TF-IDF. <https://doi.org/10.1038/srep30308>
- 45 Moriyama E. (1998). Gene length and codon usage bias in *Drosophila melanogaster*, *Saccharomyces cerevisiae* and *Escherichia coli*. <https://doi.org/10.1093/nar/26.13.3188>
- 46 Urrutia AO, Hurst LD. (2003). The Signature of Selection Mediated by Expression on Human Genes. <https://doi.org/10.1101/gr.641103>
- 47 Keeling PJ, Palmer JD. (2008). Horizontal gene transfer in eukaryotic evolution. <https://doi.org/10.1038/nrg2386>
- 48 Morel B, Kozlov AM, Stamatakis A, Szöllősi GJ. (2020). GeneRax: A Tool for Species-Tree-Aware Maximum Likelihood-Based Gene Family Tree Inference under Gene Duplication, Transfer, and Loss. <https://doi.org/10.1093/molbev/msaa141>
- 49 Szöllősi GJ, Rosikiewicz W, Boussau B, Tannier E, Daubin V. (2013). Efficient Exploration of the Space of Reconciled Gene Trees. <https://doi.org/10.1093/sysbio/syt054>
- 50 Gladyshev EA, Meselson M, Arkhipova IR. (2008). Massive Horizontal Gene Transfer in Bdelloid Rotifers. <https://doi.org/10.1126/science.1156407>
- 51 Boschetti C, Carr A, Crisp A, Eyres I, Wang-Koh Y, Lubzens E, Barraclough TG, Micklem G, Tunnacliffe A. (2012). Biochemical Diversification through Foreign Gene Expression in Bdelloid Rotifers. <https://doi.org/10.1371/journal.pgen.1003035>
- 52 Ma J, Wang S, Zhu X, Sun G, Chang G, Li L, Hu X, Zhang S, Zhou Y, Song C-P, Huang J. (2022). Major episodes of horizontal gene transfer drove the evolution of land plants. <https://doi.org/10.1016/j.molp.2022.02.001>
- 53 Freschi L, Vincent AT, Jeukens J, Emond-Rheault J-G, Kukavica-Ibrulj I, Dupont M-J, Charette SJ, Boyle B, Levesque RC. (2018). The *Pseudomonas aeruginosa* Pan-Genome Provides New Insights on Its Population Structure, Horizontal Gene Transfer, and Pathogenicity. <https://doi.org/10.1093/gbe/evy259>
- 54 Fan X, Qiu H, Han W, Wang Y, Xu D, Zhang X, Bhattacharya D, Ye N. (2020). Phytoplankton pangenome reveals extensive prokaryotic horizontal gene transfer of diverse functions. <https://doi.org/10.1126/sciadv.aba0111>

- 55 Stanhope MJ, Lupas A, Italia MJ, Koretke KK, Volker C, Brown JR. (2001). Phylogenetic analyses do not support horizontal gene transfers from bacteria to vertebrates. <https://doi.org/10.1038/35082058>
- 56 Boothby TC, Tenlen JR, Smith FW, Wang JR, Patanella KA, Osborne Nishimura E, Tintori SC, Li Q, Jones CD, Yandell M, Messina DN, Glasscock J, Goldstein B. (2015). Evidence for extensive horizontal gene transfer from the draft genome of a tardigrade. <https://doi.org/10.1073/pnas.1510461112>
- 57 Consortium IHGS, Whitehead Institute for Biomedical Research C for GR, Centre: TS, Center WUGS, Institute: UDJG, of Medicine Human Genome Sequencing Center: BC, Center: RGS, and CNRS UMR-8030: G, Department of Genome Analysis I of MB, Center: GS, Center: BGIG, Multimegabase Sequencing Center TI for SB, Center: SGT, of Oklahoma's Advanced Center for Genome Technology: U, for Molecular Genetics: MPI, Cold Spring Harbor Laboratory LAHGC, for Biotechnology: GRC, *Genome Analysis Group (listed in alphabetical order also includes individuals listed under other headings);, Scientific management: National Human Genome Research Institute UNI of H, Center: SHG, of Washington Genome Center: U, Department of Molecular Biology KUS of M, of Texas Southwestern Medical Center at Dallas: U, Office of Science UD of E, Trust: TW. (n.d.). Initial sequencing and analysis of the human genome. <https://doi.org/10.1038/35057062>
- 58 Steinegger M, Salzberg SL. (2020). Terminating contamination: large-scale search identifies more than 2,000,000 contaminated entries in GenBank. <https://doi.org/10.1186/s13059-020-02023-1>
- 59 Reiter T. (2024). Clustering the NCBI nr database to reduce database size and enable faster BLAST searches. <https://doi.org/10.57844/ARCADIA-W8XT-PC81>
- 60 Köster J, Rahmann S. (2012). Snakemake—a scalable bioinformatics workflow engine. <https://doi.org/10.1093/bioinformatics/bts480>
- 61 Di Tommaso P, Chatzou M, Floden EW, Barja PP, Palumbo E, Notredame C. (2017). Nextflow enables reproducible computational workflows. <https://doi.org/10.1038/nbt.3820>
- 62 Sayers EW, Cavanaugh M, Clark K, Pruitt KD, Schoch CL, Sherry ST, Karsch-Mizrachi I. (2021). GenBank. <https://doi.org/10.1093/nar/gkab1135>
- 63 O'Leary NA, Wright MW, Brister JR, Ciufo S, Haddad D, McVeigh R, Rajput B, Robbertse B, Smith-White B, Ako-Adjei D, Astashyn A, Badretdin A, Bao Y, Blinkova O, Brover V, Chetvernin V, Choi J, Cox E, Ermolaeva O, Farrell CM, Goldfarb T, Gupta T, Haft D, Hatcher E, Hlavina W, Joardar VS, Kodali VK, Li W, Maglott D, Masterson P, McGarvey KM, Murphy MR, O'Neill K, Pujar S, Rangwala

SH, Rausch D, Riddick LD, Schoch C, Shkeda A, Storz SS, Sun H, Thibaud-Nissen F, Tolstoy I, Tully RE, Vatsan AR, Wallin C, Webb D, Wu W, Landrum MJ, Kimchi A, Tatusova T, DiCuccio M, Kitts P, Murphy TD, Pruitt KD. (2015). Reference sequence (RefSeq) database at NCBI: current status, taxonomic expansion, and functional annotation. <https://doi.org/10.1093/nar/gkv1189>

- 64 Steinegger M, Söding J. (2017). MMseqs2 enables sensitive protein sequence searching for the analysis of massive data sets. <https://doi.org/10.1038/nbt.3988>
- 65 Rice P, Longden I, Bleasby A. (2000). EMBOSS: The European Molecular Biology Open Software Suite. [https://doi.org/10.1016/s0168-9525\(00\)02024-2](https://doi.org/10.1016/s0168-9525(00)02024-2)
- 66 Glick L, Mayrose I. (2021). Panoramic: A package for constructing eukaryotic pan-genomes. <https://doi.org/10.1111/1755-0998.13344>
- 67 Hu Z, Wei C, Li Z. (2020). Computational Strategies for Eukaryotic Pangenome Analyses. https://doi.org/10.1007/978-3-030-38281-0_13
- 68 Müllner D. (2013). **fastcluster**: Fast Hierarchical, Agglomerative Clustering Routines for *RandPython*. <https://doi.org/10.18637/jss.v053.i09>
- 69 Moura A, Savageau MA, Alves R. (2013). Relative Amino Acid Composition Signatures of Organisms and Environments. <https://doi.org/10.1371/journal.pone.0077319>
- 70 Buchfink B, Reuter K, Drost H-G. (2021). Sensitive protein alignments at tree-of-life scale using DIAMOND. <https://doi.org/10.1038/s41592-021-01101-x>
- 71 Wickham H, Averick M, Bryan J, Chang W, McGowan L, François R, Grolemund G, Hayes A, Henry L, Hester J, Kuhn M, Pedersen T, Miller E, Bache S, Müller K, Ooms J, Robinson D, Seidel D, Spinu V, Takahashi K, Vaughan D, Wilke C, Woo K, Yutani H. (2019). Welcome to the Tidyverse. <https://doi.org/10.21105/joss.01686>
- 72 Emms DM, Kelly S. (2015). OrthoFinder: solving fundamental biases in whole genome comparisons dramatically improves orthogroup inference accuracy. <https://doi.org/10.1186/s13059-015-0721-2>
- 73 Aramaki T, Blanc-Mathieu R, Endo H, Ohkubo K, Kanehisa M, Goto S, Ogata H. (2019). KofamKOALA: KEGG Ortholog assignment based on profile HMM and adaptive score threshold. <https://doi.org/10.1093/bioinformatics/btz859>
- 74 Chou S, Reiter T. (2024). Speeding up the quality control of raw sequencing data using seqqc, a Nextflow-based solution. <https://doi.org/10.57844/ARCADIA-CXN6-CH62>

- 75 Breitwieser FP, Pertea M, Zimin AV, Salzberg SL. (2019). Human contamination in bacterial genomes has created thousands of spurious proteins.
<https://doi.org/10.1101/gr.245373.118>
- 76 Crisp A, Boschetti C, Perry M, Tunnacliffe A, Micklem G. (2015). Expression of multiple horizontally acquired genes is a hallmark of both vertebrate and invertebrate genomes. <https://doi.org/10.1186/s13059-015-0607-3>
- 77 Haimlich S, Fridman Y, Khandal H, Savaldi-Goldstein S, Levy A. (2022). Widespread horizontal gene transfer between plants and their microbiota.
<https://doi.org/10.1101/2022.08.25.505314>
- 78 Salzberg SL. (2019). Next-generation genome annotation: we still struggle to get it right. <https://doi.org/10.1186/s13059-019-1715-2>
- 79 Blaxter M, Archibald JM, Childers AK, Coddington JA, Crandall KA, Di Palma F, Durbin R, Edwards SV, Graves JAM, Hackett KJ, Hall N, Jarvis ED, Johnson RN, Karlsson EK, Kress WJ, Kuraku S, Lawniczak MKN, Lindblad-Toh K, Lopez JV, Moran NA, Robinson GE, Ryder OA, Shapiro B, Soltis PS, Warnow T, Zhang G, Lewin HA. (2022). Why sequence all eukaryotes?
<https://doi.org/10.1073/pnas.2115636118>
-